

QuantiChrom™ Heme Assay Kit (DIHM-250)

Colorimetric Determination of Total Heme at 400 nm

DESCRIPTION

Heme is one important member of the porphyrin family. It is synthesized in both mitochondria and cytoplasm, and is a key prosthetic group for various essential proteins such as hemoglobin, cytochromes, catalases and peroxidases. Heme determination is widely practiced by researchers of various blood diseases.

Simple, direct and automation-ready procedures for measuring heme concentration are becoming popular in Research and Drug Discovery. BioAssay Systems' QuantiChrom™ Heme Assay Kit is based on an improved aqueous alkaline solution method, in which the heme is converted into a uniform colored form. The intensity of color, measured at 400 nm, is directly proportional to the heme concentration in the sample. The optimized formulation substantially reduces interference by substances in the raw samples and exhibits high sensitivity.

APPLICATIONS

Direct Assays: total heme in blood, serum, plasma, urine, heme-carrying enzymes.

Pharmacology: effects of drugs on heme metabolism.

Drug Discovery: HTS for drugs that modulate heme levels.

KEY FEATURES

Sensitive and accurate. Linear detection range 0.6 – 125 μM heme in 96-well plate assay.

Simple and high-throughput. The “mix-and-read” procedure involves addition of a single working reagent and reading the optical density. Can be readily automated as a high-throughput assay in 96-well plates for thousands of samples per day.

Safety. Reagents are non-toxic.

Versatility. Assays can be executed in 96-well plate or cuvet.

KIT CONTENTS (250 tests in 96-well plates)

Reagent: 50 mL

Standard: 10 mL (=125 μM heme)

Storage conditions. Store reagent and standard at 4°C. Shelf life: 12 months.

Precautions: reagents are for research use only. Normal precautions for laboratory reagents should be exercised while using the reagents. Please refer to Material Safety Data Sheet for detailed information.

PROCEDURES

Procedure using 96-well plate:

1. **Blank and Standard.** Pipette 50 μL water (Blank) and 50 μL Standard into wells of a clear bottom 96-well plate. Transfer 200 μL water into the blank and standard wells.

2. **Samples.** Serum and plasma samples can be assayed directly ($n = 1$). Blood samples should be diluted 100-fold in distilled water ($n = 100$).

Transfer 50 μL samples into wells. Important: avoid bubble formation during the pipetting steps.

3. Add 200 μL Reagent to sample wells and tap plate lightly to mix.

4. Incubate 5 min at room temperature. Read OD at 380-420nm (peak 400nm). Signal is stable for at least 2 hours.

Procedure using cuvette:

1. Transfer 100 μL sample and 1000 μL Reagent into a cuvet and tap lightly to mix. Read OD at 380-420nm (peak 400 nm) against water.

2. Transfer 100 μL Standard and 1000 μL water to cuvet. Read OD at 400nm against water.

CALCULATION

Subtract blank OD (water) from the Standard and Sample OD values. The total heme concentration of Sample is calculated as

$$= \frac{OD_{\text{SAMPLE}} - OD_{\text{BLANK}}}{OD_{\text{STD}} - OD_{\text{BLANK}}} \times 125 \times n \text{ (}\mu\text{M)}$$

OD_{SAMPLE} , OD_{STD} and OD_{BLANK} are OD values of the sample, the Standard and water. n is the dilution factor (100 for blood samples).

Conversions: 1mg/dL heme equals 15.3 μM , 0.001% or 10 ppm.

MATERIALS REQUIRED, BUT NOT PROVIDED

Pipeting devices and accessories.

Procedure using 96-well plate:

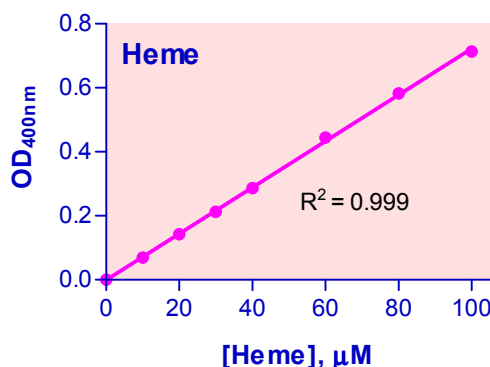
Clear-bottom 96-well plates (e.g. Corning Costar) and plate reader.

Procedure using cuvette:

Cuvets and spectrophotometer.

EXAMPLES

Heme was determined using the 96-well plate protocol. The values were $27.3 \pm 0.2 \mu\text{M}$ for rat serum, $7.8 \pm 0.4 \mu\text{M}$ for human plasma and $11.2 \pm 0.2 \text{ mM}$ for a mouse whole blood sample.



Calibration curve in 96-well plate

LITERATURE

1. Day P, Smith DW, Williams RJ (1967). Crystal spectra of a heme and some heme-protein complexes. *Biochemistry* 6:1563-1566.

2. York JL, McCoy S, Taylor DN, Caughey WS (1967). Heme A of cytochrome c oxidase. I. Isolation from bovine heart. *J Biol Chem.* 242:908-911.

3. Scholl F (1966). On the determination of heme and heme derivatives in serum. *Wien Klin Wochenschr.* 78:487.