

# AssayMax Mouse Tumor Necrosis Factor alpha (TNF-alpha) ELISA Kit

Catalog Number EMT2010-1 Lot #

#### Introduction

Tumor necrosis factor alpha (TNF-alpha) is a potent cytokine with a myriad of innate immune anti-tumor properties. TNF-alpha has a critical role in the bone and cartilage damage associated with rheumatoid arthritis (RA)[1]. TNF-alpha may be involved in the pathogenesis and/or progression of gestational diabetes mellitus (GDM) [2]. TNF-alpha is expressed in myocardium during compensated pressure-overload hypertrophy and contributes to postischemic myocardial dysfunction [3]. The serum levels of TNF-alpha were also significantly elevated in active WG (Wegener's granulomatosis) [4], in the late stages of HIV-associated disease [5], and in the spinal cord of arthritic patients [6].

## **Principal of the Assay**

The AssayMax Mouse TNF-alpha ELISA kit is designed for detection of TNF-alpha in mouse serum or cell culture supernatants. This assay employs a quantitative sandwich enzyme immunoassay technique that measures TNF-alpha in 3.5 hours. A murine monoclonal antibody specific for mouse TNF-alpha has been pre-coated onto a microplate. TNF-alpha in standards and samples is sandwiched by the immobilized antibody and a biotinylated polyclonal antibody specific for mouse TNF-alpha, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is then washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

# **Caution and Warning**

- This kit is for research use only.
- The kit should not be used beyond the expiration date.
- The Stop Solution is an acid solution.

## Reagents

- **TNF-alpha Microplate:** A 96-well polystyrene microplate (12 strips of 8 wells) coated with a murine monoclonal antibody against TNF-alpha.
- **Sealing Tapes:** Each kit contains 3 pre-cut, pressure-sensitive sealing tapes that can be cut to fit the format of the individual assay.

- **TNF-alpha Standard:** Recombinant mouse TNF-alpha in a buffered protein base (4 ng, lyophilized).
- **Biotinylated TNF-alpha Antibody (100x):** A 100-fold biotinylated polyclonal antibody against TNF-alpha (80 μl).
- Streptavidin-Peroxidase Conjugate (SP Conjugate): A 100-fold concentrate (120 µl).
- EIA Diluent Concentrate (10x): A 10-fold buffered protein base (20 ml).
- Wash Buffer Concentrate (10x): A 10-fold concentrated buffered surfactant (2 x 30 ml).
- **Chromogen Substrate**: A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- **Stop Solution**: A 0.5 N hydroxychloric acid (12 ml) to stop the chromogen substrate reaction.

## **Storage Condition**

- Store unopened kit at 2-8°C up to expiration date.
- Opened reagents may be stored for up to 1 month at 2-8°C. Store reconstituted mTNF-alpha standard at -20°C or below.
- Opened unused strip wells may return to the foil pouch with the desiccant pack, reseal along with zip-seal. May be stored for up to 1 month in a vacuum desiccator.

## **Other Supplies Required**

- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes (1-20 μl, 20-200 μl, and multiple channel)
- Deionized or distilled reagent grade water

# Sample Collection, Preparation and Storage

- **Serum:** Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 2,000x g for 10 minutes. Dilute samples 1:4 into EIA Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- Cell Culture Supernatants: Centrifuge cell culture media at 2,000x g for 10 minutes to remove debris. Collect supernatants and assay. The samples can be stored at -20°C or below. Avoid repeated freeze-thaw cycles.

# Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use. If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.
- Standard Curve: Reconstitute the 4 ng of mouse TNF-alpha Standard with 1 ml of EIA Diluent to generate a 4 ng/ml of solution. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare triplicate standard points by serially diluting the TNF-alpha standard solution twofold with equal volume of EIA Diluent to produce 2, 1, 0.5, 0.25, 0.125, and 0.0625 ng/ml. EIA Diluent serves as the zero standard (0 ng/ml).

Standard Point	Dilution	[TNF-alpha] (ng/ml)
P1	Standard (4 ng/ml)	4.000
P2	1 part P1 + 1 part EIA Diluent	2.000
P3	1 part P2 + 1 part EIA Diluent	1.000
P4	1 part P3 + 1 part EIA Diluent	0.500
P5	1 part P4 + 1 part EIA Diluent	0.250
P6	1 part P5 + 1 part EIA Diluent	0.125
P7	1 part P6 + 1 part EIA Diluent	0.063
P8	EIA Diluent	0.000

- **EIA Diluent Concentrate** (10x): Dilute the EIA Diluent Concentrate 1:10 with reagent grade water.
- **Biotinylated TNF-alpha Antibody (100x):** Spin down the antibody briefly and dilute the desired amount of the antibody 1:100 with EIA Diluent.
- Wash Buffer Concentrate (10x): Dilute the Wash Buffer Concentrate 1:10 with reagent grade water.
- **SP Conjugate (100x):** Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with EIA Diluent.

#### **Assay Procedure**

- Prepare all reagents, working standards and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-30°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil
  pouch with desiccant inside. Reseal the pouch securely to minimize exposure to water vapor
  and store in a vacuum desiccator.
- Add  $50 \mu l$  of Standard or sample per well. Cover wells and incubate for two hours. Start the timer after the last sample addition.
- Wash five times with 200 µl of Wash Buffer. Invert the plate and decant the contents, and hit it 4-5 times on absorbent paper towel to complete remove liquid at each step.
- Add 50 μl of Biotinylated TNF-alpha Antibody to each well and incubate for 60 minutes.
- Wash five times with 200 µl of Wash Buffer as above.
- Add 50 µl of Streptavidin-Peroxidase Conjugate per well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash five times with 200 µl of Wash Buffer as above.
- Add 50 μl of Chromogen Substrate per well and incubate for approximately 10 minutes or till the optimal blue color density develops. Gently tap the plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 µl of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm **immediately**. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

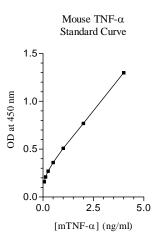
# **Data Analysis**

- Calculate the mean value of the triplicate readings for each standard and sample.
- To generate a Standard Curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis of the linear portion of the curve.

• Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

#### **Standard Curve**

• The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.



## **Sensitivity and Specificity**

- The minimum detectable dose of TNF-alpha is typically < 15 pg/ml.
- Intra-assay and inter-assay coefficients of variation were 5.5% and 8.2% respectively.
- This assay recognizes both natural and recombinant mouse TNF-alpha.

#### References

- 1. Taylor PC. (2001) Mol. Biotechnol. 19(2): 153-68
- 2. Coughlan MT et al. (2001) Diabet. Med. 18(11): 921-7
- 3. Stamm C et al. (2001) Circulation 104(12 Suppl 1): I350-5
- 4. Ohta N et al. (2001) Auris. Nasus. Larynx. 28(4): 311-4
- 5. Caso G et al. (2001) Clin. Sci. (Lond) 101(6): 583-9
- 6. Nanki T et al. (2001) J. Immunol. 167(9): 5381-5

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