
Product Manual

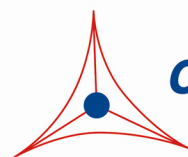
CytoSelect™ 24-Well Cell Migration Assay (8 μ m, Colorimetric Format)

Catalog Number

CBA-100

12 assays

FOR RESEARCH USE ONLY
Not for use in diagnostic procedures



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Creating Solutions for Life Science Research

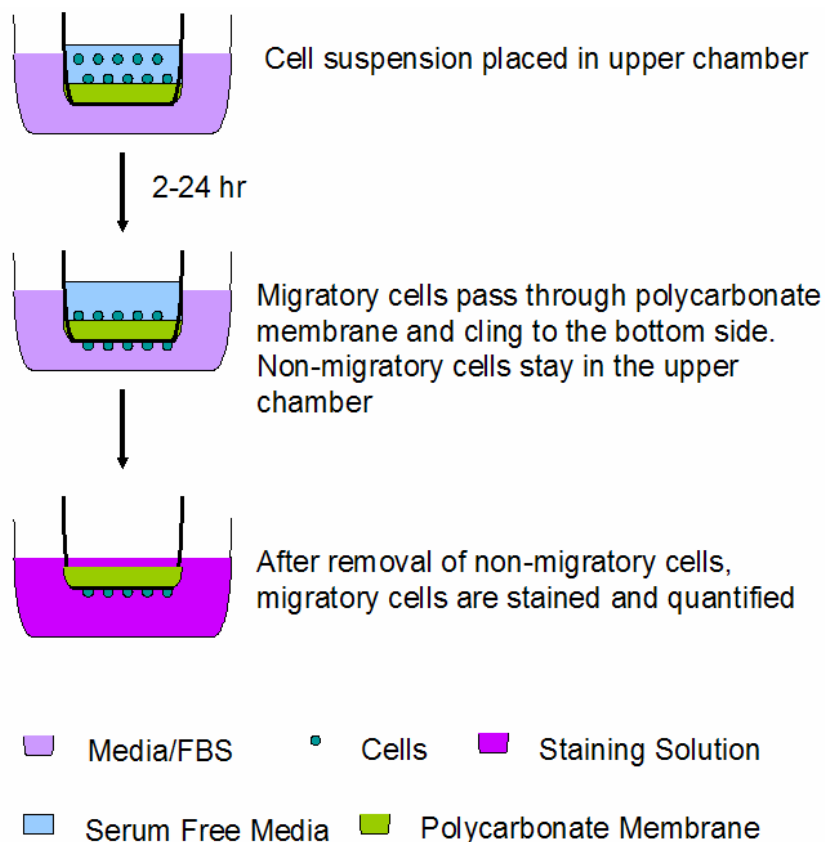
Introduction

Cell migration is a highly integrated, multistep process that orchestrates embryonic morphogenesis, tissue repair and regeneration. It plays a pivotal role in the disease progression of cancer, mental retardation, atherosclerosis, and arthritis. The initial response of a cell to a migration-promoting agent is to polarize and extend protrusions in the direction of the attractant; these protrusions can consist of large, broad lamellipodia or spike-like filopodia. In either case, these protrusions are driven by actin polymerization and can be stabilized by extracellular matrix (ECM) adhesion or cell-cell interactions (via transmembrane receptors).

Cell Biolabs CytoSelect™ Cell Migration Assay Kit utilizes polycarbonate membrane inserts (8 µm pore size) to assay the migratory properties of cells. The kit contains sufficient reagents for the evaluation of 12 samples. The 8 µm pore size is optimal for epithelial and fibroblast cell migration. However, in the case of leukocyte chemotaxis, a smaller pore size (3 µm) is recommended.

Assay Principle

The CytoSelect™ Cell Migration Assay Kit contains polycarbonate membrane inserts (8 µm pore size) in a 24-well plate. The membrane serves as a barrier to discriminate migratory cells from non-migratory cells. Migratory cells are able to extend protrusions towards chemoattractants (via actin cytoskeleton reorganization) and ultimately pass through the pores of the polycarbonate membrane. Finally, the cells are removed from the top of the membrane and the migratory cells are stained and quantified.



Related Products

1. CBA-100-C: CytoSelect™ 24-Well Cell Migration and Invasion Assay (8µm, Colorimetric)
2. CBA-100-COL: CytoSelect™ 24-Well Cell Haptotaxis Assay (Collagen I, Colorimetric)
3. CBA-100-FN: CytoSelect™ 24-Well Cell Haptotaxis Assay (Fibronectin, Colorimetric)
4. CBA-101: CytoSelect™ 24-Well Cell Migration Assay (8µm, Fluorometric)
5. CBA-102: CytoSelect™ 24-Well Cell Migration Assay (5µm, Fluorometric)
6. CBA-103: CytoSelect™ 24-Well Cell Migration Assay (3µm, Fluorometric)
7. CBA-106: CytoSelect™ 96-Well Cell Migration Assay (8µm, Fluorometric)
8. CBA-110: CytoSelect™ 24-Well Cell Invasion Assay (Basement Membrane, Colorimetric)
9. CBA-130: CytoSelect™ 96-Well Cell Transformation Assay (Soft Agar Colony Formation)

Kit Components

1. 24-well Migration Plate (Part No. 10001): One 24-well plate containing 12 cell culture inserts (8 µm pore size)
2. Cell Stain Solution (Part No. 11002): One Bottle – 10.0 mL
3. Extraction Solution (Part No. 11003): One Bottle – 10.0 mL
4. Cotton Swabs (Part No. 11004): 40 each.
5. Forceps (Part No. 11005): One each.

Materials Not Supplied

1. Migratory cell lines
2. Cell culture medium
3. Serum free medium, such as DMEM containing 0.5% BSA, 2 mM CaCl₂ and 2 mM MgCl₂
4. Cell culture incubator (37 °C, 5% CO₂ atmosphere)
5. Light microscope
6. 96-well microtiter plate
7. Microtiter plate reader

Storage

Store all components at 4°C until their expiration dates.

Assay Protocol

1. Under sterile conditions, allow the 24-well migration plate to warm up at room temperature for 10 minutes.
2. Prepare a cell suspension containing $0.5\text{-}1.0 \times 10^6$ cells/ml in serum free media. Agents that inhibit or stimulate cell migration can be added directly to the cell suspension.
Note: Overnight starvation may be performed prior to running the assay
3. Add 500 μL of media containing 10% fetal bovine serum or desired chemoattractant(s) to the lower well of the migration plate.
4. Add 300 μL of the cell suspension solution to the inside of each insert.
5. Incubate for 2-24 hours in a cell culture incubator.
6. Carefully aspirate the media from the inside of the insert. Wet the ends of 2-3 cotton-tipped swabs and gently swab the interior of the inserts to remove non-migratory cells. Take care not to puncture the polycarbonate membrane. Be sure to remove cells on the inside perimeter of the insert.
7. Transfer the insert to a clean well containing 400 μL of Cell Stain Solution and incubate for 10 minutes at room temperature.
8. Gently wash the stained inserts several times in a beaker of water. Allow the inserts to air dry.
9. (optional) Count migratory cells with a light microscope under high magnification objective, with at least three individual fields per insert.
10. Transfer each insert to an empty well, adding 200 μL of Extraction Solution per well, then incubating 10 minutes on an orbital shaker.
11. Transfer 100 μL from each sample to a 96-well microtiter plate and measure the OD 560nm in a plate reader.

Example of Results

The following figures demonstrate typical with the CytoSelect™ Cell Migration Assay Kit. One should use the data below for reference only. This data should not be used to interpret actual results.

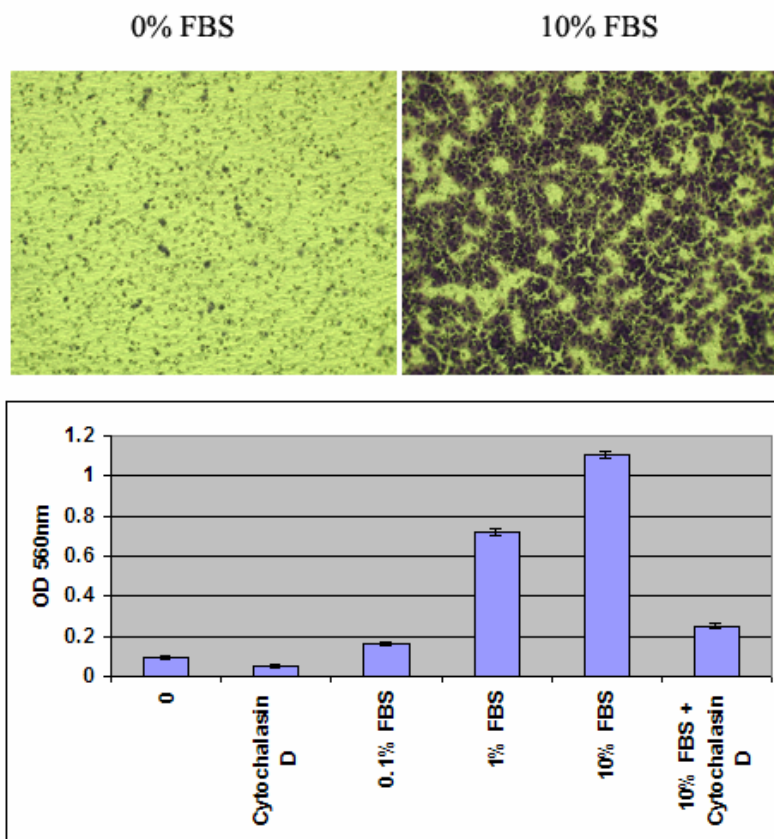


Figure 1. Human Fibrosarcoma HT-1080 Cell Migration. HT-1080 cells were seeded at 150,000 cells/well and allowed to migrate toward FBS for 4 hrs in the presence or absence of 2 μ M Cytochalasin D. Migratory cells on the bottom of the polycarbonate membrane were stained (top panel picture) and quantified at OD 560nm after extraction (bottom panel figure).

References

1. Ridley AJ, Schwartz MA, Burridge K, Firtel RA, Ginsberg MH, Borisy G, Parsons JT, Horwitz AR. (2003) *Science* **302**, 1704-9.
2. Horwitz R, Webb D. (2003) *Curr Biol.* **13**, R756-9.
3. Lauffenburger DA, Horwitz AF. (1996) *Cell* **84**, 359-369.

Recent Product Citations

1. Murtagh, J. et al. (2006). Taxotere-induced inhibition of human endothelial cell migration is a result of Heat Shock Protein 90 degradation. *Cancer Research* **66**: 8192-8199.
2. Lu, H. et al. (2006). The microtubule binding drug laulimalide inhibits Vascular Endothelial Growth Factor-induced human endothelial cell migration and is synergistic when combined with Docetaxel (Taxotere). *Mol Pharmacol.* **69(4)**: 1207-1215.
3. Phinney, D. et al. (2006). Murine mesenchymal stem cells transplanted to the central nervous system of neonatal versus adult mice exhibit distinct engraftment kinetics and express receptors that guide neuronal cell migration. *Stem Cells Dev.* **15(3)**: 437-447.
4. Isakova, I. et al. (2007). Age and dose-related effects on MSC engraftment levels and anatomical distribution in the CNS of non-human primates: Identification of novel MSC subpopulations that respond to guidance cues in brain. *Stem Cells* **25**: 3261-3270.
5. Eckstein, N. et al. (2008). EGFR-pathway analysis identifies amphiregulin as a key factor for cisplatin resistance of human breast cancer cells. *J. Biol. Chem.* **283**: 739-750.
6. Liu, S. et al. (2008). Matrix metalloproteinase-activated anthrax lethal toxin demonstrates high potency in targeting tumor vasculature. *J. Biol. Chem.* **283**: 529-540.

Warranty

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